

Early cranial nerve dysfunction is correlated to altered facial morphology in spinocerebellar ataxia type 2. Cranial nerves and facial morphology in SCA2

Disfunción temprana de nervios craneales correlaciona con alteraciones de la morfología facial en la ataxia espinocerebelosa tipo 2.

Los nervios craneales y la morfología facial en SCA2

Jacqueline Medrano-Montero ^{1,2}, Luis Velázquez-Pérez ^{1,2}, Roberto Rodríguez-Labrada ^{1,2}, Nalia Canales-Ochoa ^{1,2}, Arnoy Peña-Acosta ^{1,2}, Luis E Almaguer-Mederos ^{1,3}, Annelies Estupiñán-Rodríguez ^{1,2}, Georg Auburger ⁴.

Abstract

The aim of our cross-sectional study was to quantify trigeminal and facial nerve electrophysiological alterations and to determine its correlation with facial morphology abnormalities and expanded CAG repeats in Spinocerebellar ataxia type 2 (SCA2). 90 SCA2 patients and 41 preclinical mutation carriers together with 100 sex-, age- and facial type- matched healthy subjects as controls were assessed by facial motor nerve conduction, blink reflex (BR) and mandibular reflex (jaw jerk). Facial morphology features were analyzed by the determination of the facial type using a standardized morphometric facial index and the measurement of three distinct planes over pictures. Patients exhibited a significant prolongation of latency and duration and decreased amplitude in the facial motor potentials. The mandibular reflex revealed prolonged latency and decreased amplitude. Moreover, the bilateral R2 component of the blink reflex was prolonged. Preclinical carriers showed prolonged duration for facial nerve potentials and mandibular reflex, as well as increased latency of bilateral R2 BR component. Facial morphology measures revealed periorbital, perioral and masseter alterations in patient and preclinical groups, and some of them were correlated to the electrophysiological features and expanded CAG repeats.

These electrophysiological and morphological features widen the prodromal phenotype of SCA2, and offer new clues about the role of ATXN2 mutations for muscle atrophy, neuronal energy balance and lipid metabolism.

Resumen:

Se realizó un estudio transversal con el objetivo de cuantificar las alteraciones electrofisiológicas de los nervios facial y trigémino y determinar su correlación con las anomalías de la morfología facial y el número de repeticiones de CAG en la Ataxia Espinocerebelosa Tipo 2 (SCA2). Se evaluaron 90 pacientes SCA2 y 41 portadores preclínicos de mutación junto con 100 sujetos sanos como controles, pareados por sexo, edad y tipo facial a través de estudios de conducción nerviosa motora periférica del nervio facial, reflejo de parpadeo (BR) y reflejo T mentoniano (reflejo mandibular). Para el análisis de las características de la morfología facial se determinó el tipo facial mediante el índice facial morfológico estandarizado y mediciones a partir de tres planos distintos sobre fotografías. Los pacientes mostraron una prolongación significativa de la latencia y la duración y una reducción de la amplitud del potencial motor del facial. El reflejo mandibular reveló prolongación de la latencia y disminución de la amplitud así como prolongación del componente R2 bilateral del BR. Los portadores preclínicos mostraron duración prolongada del potencial del nervio facial, del reflejo mandibular y de la latencia del componente R2 bilateral del BR. Se obtuvieron alteraciones morfológicas faciales sobre los músculos periorales, periorbitarios y maseterinos en ambos grupos, y algunas de ellas correlacionaron con las hallazgos electrofisiológicos y el número de repeticiones de CAG. Estas características electrofisiológicas y morfológicas amplían el fenotipo prodrómico de SCA2 y ofrecen nuevas pistas sobre el papel de la mutación ATXN2 en la atrofia muscular, el equilibrio energético neuronal y el metabolismo lipídico.

¹ Clinic

² Dept. Clinical Neurophysiology, Center for Research and Rehabilitation of the Hereditary Ataxias (CIRAH), Libertad Street 26, Holguín 80100, Cuba.

³ Dept. Molecular Neurobiology, Center for Research and Rehabilitation of the Hereditary Ataxias (CIRAH), Libertad Street 26, Holguín 80100, Cuba.

⁴ Exp. Neurology, Goethe University Medical School, 60590 Frankfurt am Main, Germany

Mailig address:

Jacqueline Medrano-Montero: Center for Research and Rehabilitation of Hereditary Ataxias (CIRAH), Libertad Street 26, Holguín 80100, Cuba. Email address: jacobita64@gmail.com, tel: +53 24422491/ 24462823.

Received: 14 de October 2017

Accepted: 11 de March 2018

Conflict of interest: Declares that any conflict of interest with any of the authors there is

Key words:

SCA2; Motor performance; hereditary ataxia; spinocerebellar ataxia type 2; olivo-ponto-cerebellar atrophy

Palabras clave:

SCA2; Funcionamiento del motor; ataxia hereditaria; ataxia espinocerebelosa tipo 2; atrofia olivo-ponto-cerebelosa

Introduction

Spinocerebellar Ataxia type 2 (SCA2) is an autosomal dominant cerebellar ataxia caused by CAG repeat expansions in the ATXN2 gene on chromosome 12q¹⁻⁶. It is characterized by several core symptoms including a progressive cerebellar ataxia with dysarthria, slowing of horizontal saccades in more than 90% of cases, peripheral neuropathy, dysphagia, olfactory deterioration, autonomic abnormalities, sleep disturbances and cognitive dysfunction leading to reduced survival⁷⁻¹⁶. In Cuba, there are about 578 living SCA2 patients from 163 families and 7,200 asymptomatic at-risk individuals, representing an estimated ATXN2 mutation prevalence of nearly 28.5 cases per 100 000 inhabitants in the whole country. SCA2 represents 87% of all SCA subtypes in Cuba. In Holguin province, the prevalence rate is about 40.18 cases per 100 000 inhabitants, distributed in ten municipalities of the province. This topographic location gives unique characteristics to the disease in our country and is supposed to be considered a founder gene effect^{9,17}.

Although most phenotypical features of SCA2 have been characterized thoroughly with clinical, electrophysiological and neuropathological approaches^{7, 8,10-12, 14,17-31}, the prominent affection of the cranio-cervical area during the preclinical and early disease stages deserve special attention, particularly the morphological alterations of the face. It has been known for some time that SCA2 and SCA3 patients within the first years of disease appear distracted due to their characteristic belated gaze reaction, that they show fasciculation-like movements in a cranio-cervical distribution and seem to be frightened or have a mask-like face. This is due to additional lid retraction, probably a product of subcutaneous fat loss, autonomic pathology and cranial nerve degeneration^{3, 19,32-34}. Some parents of affected individuals have reported that they recognize the onset of pathology among their offspring first through the changes of facial morphology. However, there is no objective quantitative characterization of facial alterations and their correlation with clinical and molecular variables so far.

Material and Methods

Here we performed a cross-sectional assessment of facial alterations by neurophysiological studies and morphological measurements and in 90 SCA2 patients (37 female and 53 male subjects), 41 pre-clinical carriers (28 female and 13 male) and 100 healthy subjects (60 female, 40 male) as controls.

Inclusion criteria for SCA2 patients were: i) age between 15 and 65 years old, ii) positive molecular diagnosis of SCA2 mutation and iii) presence of definite cerebellar syndrome. In the case of pre-clinical carriers, the absence of definite cerebellar syndrome was assumed as third inclusion criteria. The control group was selected following these criteria: i) absence of cerebellar manifestations by exhaustive neurological examination, ii) absence of ataxia or other related diseases in the family history, and iii) age, gender and facial type matching with patients and preclinical carriers.

Exclusion criteria for all participants were: chronic alcohol abuse or use of CNS- depressant drugs, psychiatric disorders and patients with other diseases affecting the nervous system such as hypothyroidism, diabetes, hypertension and others. Also, subjects with history of facial trauma were not included.

The study protocol was approved by the ethical standards of the committee on human experimentation of the Centre for Research and Rehabilitation of Hereditary Ataxias and was in agreement with the Helsinki declaration. All participants gave their written informed consent prior to the experiments.

Neurological assessments

All subjects underwent a complete neurological examination and the International Cooperative Ataxia Rating Scale (ICARS) to assess the severity of the cerebellar syndrome³⁵.

Neurophysiological studies

Neurophysiological evaluations were also performed in all enrolled subjects and consisted in facial motor nerve conduction, blink reflex (BR) and mandibular reflex (jaw jerk, JJ) studies, which were conducted using standardized protocols³⁶ by a researcher blinded to the experimental group of each individual.

Facial morphological measures

As previously described, human faces show considerable variance of height and other features in comparison to facial width being constant³⁷. Therefore, facial morphology analyses were conducted in 41 SCA2 patients, 41 preclinical carriers and 41 sex-, age- and facial type- matched controls. In all groups, the gender distribution was 28 females/12 males. Facial measures were conducted to classify the facial type and to quantify the morphological alterations. The determination of the facial type was performed by direct measures with a cranio

meter using a morphologic facial index³⁸. Accordingly, three facial types were defined: i) Leptoprosopo or long face (index $\frac{\text{length}}{\text{width}} > 104$), Mesoprosopo or intermediate face (index 97-104) and Euriprosopo or wide face (index < 97).

Frontal and right view photography to each subject were taken, with Kodak DC290 Zoom digital camera (November 1999, Canada), with head orientation in Frankfort plane parallel to the floor and 1m focal length.

Measures to quantify the facial alterations were performed indirectly on frontal and lateral right view pictures using Adobe Photoshop program version 7. Measures were assessed using the following standard landmarks and lines as references: Frankfort (FP), Mid Sagittal (MSP) and commissural (CP) planes for frontal views and FP, anterior frontal (Izard) and later frontal (Simon) planes for lateral right views (Fig. 1). For the frontal views the measures were assessed bilaterally and consisted in:

- (1) Distance from MSP to internal eye angle (MSP-IEA).
- (2) Distance from MSP to external eye angle (MSP-EEA)
- (3) Distance from MSP to the more depressed point of the cheek (MSP-DPC)
- (4) Distance from MSP to the labial commissure of both sides in the CP (MSP-LC)
- (5) Distance from FP to labial commissure (perpendicular) (FP-LC)
- (6) Distance from FP to internal eye angle (FP-IEA)
- (7) Distance from FP to external eye angle (FP-EEA)

In the lateral right views measures were:

- (1) Distance from FP to labial commissure (perpendicular) (FP-LC),
- (2) Distance from Simon plane to labial commissure (SP-LC),
- (3) Distance from Izard plane to the more depressed point of the upper eyelid (IP-DPUE),
- (4) Distance from Izard plane to the more depressed point of the lower eyelid (IP-DPLE).

All morphological measures were conducted by a researcher blinded to the experimental group of each individual

CAG repeat quantification

The ATXN2 CAG repeat length was assessed by PCR amplification followed by polyacrylamide gel electrophoresis³⁹.

Lateral right view pictures, to quantify the facial alterations.

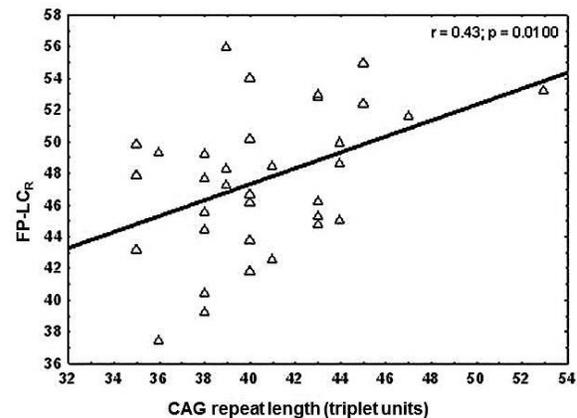


Figure 3. Correlation between CAG repeat length and the morphometric distance FP-LCR.

Statistical Analysis

Collected information from this study was entered into a data file using the Statistic for Windows (v 6.1, 2003, USA) computer software. Scatter plots were obtained to evaluate the data and to confirm the presence of a normal distribution before the application of parametric statistics. Descriptive statistics of the studied variables, analyses of univariate variance, and Spearman's correlation tests between electrophysiological, morphologic and molecular variables were performed.

Results

General characteristics of study population

All patients had clinical and molecular diagnosis of SCA2, with age ranging between 16 to 60 years (Mean = 39.9 years; SD: 11.41), whereas age at onset varied from 8 to 60 years (Mean = 26.7 years; SD: 9.86). Disease duration was between 1 to 42 years (Mean=13.25; SD: 7.61); ICARS score mean was 50.06 (SD: 17.32) and varied from 5.88 to 92 points. Pathological CAG repeat lengths covered sizes from 34 to 53 (Mean=40.58; SD: 3.37). Normal alleles ranged between 19 and 33 CAG (Mean=22.08; SD: 1.31).

Preclinical carriers group had mean age of 36.7 years (SD: 10.27), ranging from 19 to 62 years. Pathological CAG repeat length varied between sizes 32 and 42 (Mean=36.68; SD: 2.47) and normal alleles mean was at 22.74 (SD: 2.33).

Mean age of the control group was 34.02 years (SD: 8.43; range: 20-60).

Neurophysiological findings

Findings of the mean comparison tests for neurophysiological assessments are shown in Table 1. The study of the facial (F) motor nerve conduction revealed a significant prolongation of latency ($p=0.000$) and duration ($p=0.000$) of the potential as well as a significant ($p=0.029$) decrease of the amplitude in SCA2 patients compared to controls. There was also a significant prolongation of duration ($p=0.004$) of the potential in preclinical mutation carriers (Table 1).

stage and it's progressing to axonal damage on manifest SCA2 subjects.

Facial morphometric analyses

Given that the early vulnerability of cranial nerves will also result in tissue atrophy over time, quantitative morphological assessments of facial features were attempted by standardized digital procedures. Consistent with the literature, the mesoprosopo facial type was most common among all groups. Relevant alterations of facial morphology in SCA2 patients were located (see Fig. 2) at linear distan-

Table 1. Mean comparison of neurophysiological variables in SCA2 patients, preclinical mutation carriers and control groups.

Variables Mean \pm SD	Patients	Control	Preclinical carriers	Control
Latency F (ms)	3.06 \pm 0.57 ^{***}	2.64 \pm 0.49	2.82 \pm 0.35 ^{ns}	2.64 \pm 0.05
Duration F (ms)	12.35 \pm 3.55 ^{***}	10.42 \pm 3.62	12.87 \pm 3.79 ^{**}	10.64 \pm 3.16
Amplitude F (mV)	1.82 \pm 0.78 ^{**}	2.12 \pm 1.09	1.96 \pm 0.62 ^{ns}	2.28 \pm 1.80
Latency JJ (ms)	10.35 \pm 5.60 ^{***}	5.73 \pm 1.05	8.14 \pm 2.16 ^{***}	5.85 \pm 0.97
Amplitude JJ (mV)	0.51 \pm 0.36 ^{***}	1.26 \pm 1.06	1.51 \pm 2.16 ^{ns}	1.22 \pm 0.41
R2 Ipsi. BR (ms)	37.63 \pm 5.58 ^{***}	32.38 \pm 3.38	34.31 \pm 5.01 ^{**}	32.12 \pm 3.30
R2 Contra. BR (ms)	39.62 \pm 6.63 ^{***}	33.27 \pm 3.58	35.21 \pm 5.41 ^{**}	32.67 \pm 3.34

F: facial; JJ: jaw jerk; BR: Blink reflex. Lat.: Latency; Ipsi: Ipsilateral; Contra: Contralateral. ns: next to the mean (SD) value represents no statistical differences ($p>0.05$); * means statistical difference ($p<0.05$); ** means higher statistical difference ($p<0.005$) and *** means highest statistical difference ($p<0.0005$).

In the jaw jerk (JJ) study, an increment of latency values ($p=0.000$) with a decrease of the amplitude ($p=0.000$) was observed in patients, whereas in the preclinical carriers only the prolongation of latency ($p=0.000$) was observed (Table 1).

Blink reflex (BR) alterations mainly consisted in the prolongation ($p=0.000$) of the ipsilateral and contralateral latency of the R2 component in SCA2 patients and preclinical mutation carriers compared to the control group ($p=0.020$ and $p=0.011$, respectively) (Table 1). These findings confirm the early demyelination of cranial nerves at the preclinical

stages that point to a depression of the cheek (MSP-DPCL; MSP-DPCR) ($p=0.000$) and eyelids atrophy (IP-DPUE; IP-DPLE) ($p=0.022$ and $p=0.040$, respectively). Importantly, the alterations located at distances pointing to a decline of labial commissures (FP-LCL; FP-LCR) were significant not only in SCA2 patients ($p=0.016$ and $p=0.018$, respectively), but already in the preclinical carriers ($p=0.017$ and $p=0.013$, respectively) (Table 2). These results represent the first morphometric identification of SCA2 facial shape anomalies and demonstrate the preferential affection of lip

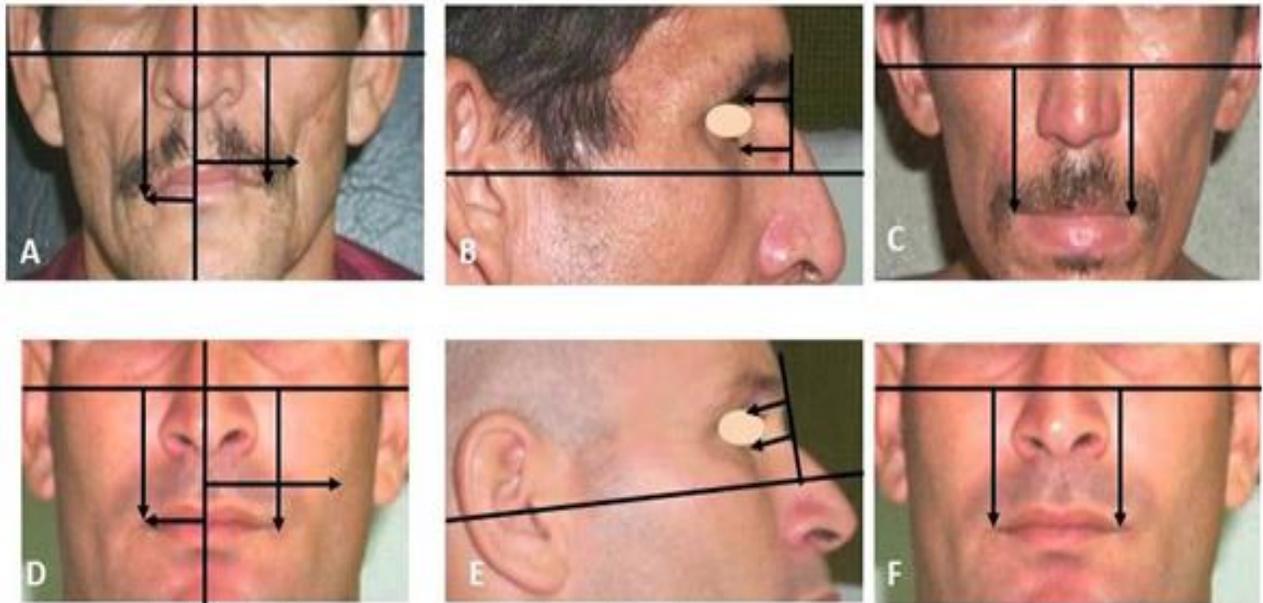


Figure 2. Indirect facial measurements on frontal and lateral right view pictures in three mesoprosopo facial type subjects. A: frontal view of a 39-years-old SCA2 patient; B: lateral right view of same SCA2 patient; C: frontal view of preclinical 34-years-old mutations carrier; D, E and F: frontal and lateral views of a healthy subject aged 34 years, as a comparison to the facial alterations above, which are illustrated by arrows in each case. (All subjects gave the consent to publish their pictures)

Table 2. Mean comparison of facial morphological measurements in SCA2 patients, preclinical carriers and control individuals

Variables (mm) Mean \pm SD	Patients	Controls	Preclinical carriers	Controls
MSP-DPC _L	41.67 \pm 4.67 ^{***}	46.32 \pm 5.13	46.51 \pm 4.64 ^{ns}	46.08 \pm 4.86
MSP-DPC _R	41.81 \pm 3.89 ^{***}	46.24 \pm 4.76	45.42 \pm 5.29 ^{ns}	45.88 \pm 4.51
MSP-LC _L	22.09 \pm 2.73 ^{**}	23.41 \pm 2.04	23.01 \pm 2.76 ^{ns}	23.41 \pm 2.17
MSP-LC _R	22.46 \pm 2.48 ^{**}	23.75 \pm 2.26	23.01 \pm 2.46 ^{ns}	24.04 \pm 2.27
FP-LC _L	47.59 \pm 4.94 ^{**}	46.64 \pm 5.12	49.41 \pm 5.02 [*]	46.65 \pm 5.13
FP-LC _R	47.22 \pm 4.90 ^{**}	46.43 \pm 4.99	49.23 \pm 4.86 [*]	46.43 \pm 4.99
IP-DPUE	14.05 \pm 3.80 [*]	12.21 \pm 3.24	11.81 \pm 3.61 ^{ns}	12.56 \pm 3.14
IP-DPLE	16.48 \pm 4.07 [*]	15.22 \pm 3.32	15.62 \pm 3.54 ^{ns}	15.27 \pm 3.30

*Morphological measurements on soft tissue. ns: next to the mean (SD) value represents no statistical differences ($p > 0.05$); * means statistical difference ($p < 0.05$); ** means higher statistical difference ($p < 0.005$) and *** means highest statistical difference ($p < 0.0005$).*

MSP-DPCL/MSP-DPCR: Midsagittal plane distance to the most depressed left/right cheek point; MSP-LCL/ MSP-LCR: Midsagittal plane distance to left/right labial commissure; FP-LCL/FP-LCR: Frankfurt plane distance to left/right labial commissure and IP-DPUE/IP-DPLE: Izard plane distance to the most depressed upper/lower eyelid point.

commissure measures during the preclinical stage of SCA2.

Factorial ANOVAs using facial measures as dependent variable and the group (patients Vs. pre-clinical carriers) and facial types (Mesoprosopo, Leptoprosopo and Euriprosopo) as factors, showed no significant interaction between the group and the facial type for none facial measure (Table 3).

Genotype-phenotype correlation analyses

area. The distance pointing to a decline of labial commissures of both sides showed positive correlations with the duration of facial potential (FP-LCL $p=0.024$; FP-LCR $p=0.014$) in SCA2 patients (Table 4A), suggesting that the pathology of the facial nerve is mirrored by the position of labial angles. The most depressed point of the superior and inferior eyelid showed significant positive correlations with ipsilateral (IP-DPUE $p=0.027$; IP-DPLE $p=0.005$) and contralateral R2 components

Table 3. Factorial ANOVAs findings for the assessment of the group x facial types interaction effect on morphometric variables

Variables (mm)	F	p
MSP-DPC _L	0.916	0.405
MSP-DPC _R	0.715	0.493
MSP-LC _L	0.185	0.831
MSP-LC _R	0.206	0.814
FP-LC _L	0.121	0.886
FP-LC _R	0.004	0.996
IP-DPUE	10.48	0.234
IP-DPLE	10.778	0.176

MSP-DPCL/MSP-DPCR: Midsagittal plane distance to the most depressed left/right cheek point; MSP-LCL/ MSP-LCR: Midsagittal plane distance to left/right labial commissure; FP-LCL/FP-LCR: Frankfurt plane distance to left/right labial commissure and IP-DPUE/IP-DPLE: Iazard plane distance to the most depressed upper/lower eyelid point

To evaluate if the electrophysiological and morphometric findings correlate with disease severity as represented by the ATXN2 CAG repeat length, and to identify the features that are particularly susceptible even at mildest stages of SCA2, we performed genotype-phenotype and electrophysiology-morphology association analyses. In SCA2 patients, Spearman’s correlation tests revealed that the CAG repeat length correlated positively with the electrophysiological latency of the facial motor nerve ($r=0.316$; $p=0.016$) and with the morphometric distance between Frankfurt plane to labial commissure ($r=0.439$; $p=0.010$) (Fig. 3). The morphometric measures that point to a sinking of the cheek (MSP-DPCL; MSP-DPCR) were inversely correlated to the mandibular reflex latency ($p=0.049$ and $p=0.039$ respectively) and directly correlated to the mandibular reflex amplitude ($p=0.003$ and $p=0.017$, respectively) in SCA2 patients (Table 4A), suggesting that this morphometric feature might reflect atrophy of the masseteric

of the BR (IP-DPUE $p=0.045$; IP-DPLE $p=0.006$) in SCA2 patients (Table 4A). In addition, the same morphometric features exhibited significant positive correlations with the facial motor nerve latency (IP-DPUE: $p=0.045$; IP-DPUE: $p=0.034$) in SCA2 patients (Table 4A), suggesting that the loss of myelin in the facial motor nerve is mirrored by the enophthalmos.

In the preclinical mutation carriers the distance between mid-sagittal plane to left labial corner was negatively correlated with the JJ reflex latency (MSP-LCL: $p=0.024$) and latency of the facial motor (MSP-LCR: $p=0,008$; MSP-LCL: $p=0.000$). The latency of the facial motor nerve also had a significant direct correlation with the measure that reflected the vertical decline of lip corners (FP-LCL: $p=0.045$; FP-LCR: $p=0.046$) (Table 4B). Thus, in preclinical carriers the demyelination of the facial motor nerve as well as lip depression were particularly conspicuous markers of incipient SCA2.

A

Table 4. Spearman's correlation test between electrophysiological and morphological variables in SCA2 patients (A) and preclinical mutation carriers (B).

Variables	Mean comparison		Spearman's correlation coefficients									
	Patients	Controls	Facial motor nerve conduction study		Blink reflex			Jaw Jerk				
			Latency	Duration	R1 Lat 1	R1 Pico	R2 Ipsi	R2 Contra	Latency	Amplitude		
MSP-DPC _L	41,67 (±4,67)	46,32 (±5,13)	- 0,055	- 0,090	- 0,069	- 0,079	- 0,005	0,002	- 0,229 *	0,342 **		
MSP-DPC _R	41,81 (±3,89)	46,24 (±4,76)	- 0,129	- 0,164	- 0,162	- 0,150	- 0,108	- 0,089	- 0,241 *	0,277 **		
FP-LC _L	47,59 (±4,99)	46,64 (±5,12)	0,156	0,262 *	0,010	0,117	0,192	0,156	0,024	0,181		
FP-LC _R	47,22 (±4,90)	46,43 (±4,99)	0,139	0,284 *	0,136	0,153	0,227	0,195	0,026	0,161		
IP-DPUE	14,05 (±3,80)	12,21 (±3,24)	0,234 *	0,160	0,077	0,123	0,256 *	0,234 *	0,216	- 0,038		
IP-DPLE	17,48 (±4,37)	15,22 (±3,32)	0,296 *	0,128	0,141	0,210	0,323 **	0,314 **	0,117	- 0,016		

B

Table 4. Spearman's correlation test between electrophysiological and morphological variables in SCA2 preclinical mutation carriers (B).

Variables	Mean comparison		Spearman's correlation coefficients									
	Preclinical carriers	Controls	Facial motor nerve conduction study		Blink reflex			Jaw Jerk				
			Latency	Duration	R1 Lat 1	R1 Pico	R2 Ipsi	R2 Contra	Latency	Amplitude		
MSP-LC _L	23,01 (±2,17)	23,41(±2,00)	- 0,420 **	0,074	0,085	- 0,324	- 0,052	- 0,024	- 0,362*	- 0,224		
MSP-LC _R	23,01 (±2,27)	23,83 (±2,17)	- 0,533 ***	- 0,022	0,182	- 0,043	- 0,183	- 0,146	- 0,278	- 0,112		
FP-LC _L	49,41 (±5,13)	46,64 (±5,12)	0,312 *	- 0,072	- 0,106	- 0,031	- 0,019	0,004	- 0,002	- 0,028		
FP-LC _R	49,23 (±4,99)	46,43 (±4,99)	0,321 *	- 0,088	- 0,114	- 0,007	- 0,024	- 0,016	0,034	- 0,020		

Legend: p ≤ 0.05: *; p ≤ 0.005: **; p ≤ 0.0005: ***

Discussion

In the present paper, we presented the first study assessing the neurophysiological abnormalities of cranial nerves and their relationship with facial morphological alterations and expanded CAG repeats in SCA2. The main findings were the presence of electrophysiological signs of myelin and axon damage within the trigeminal and facial nerves in patients and myelin lesion, predominantly in small caliber facial's fibres on preclinical mutation carriers, which correlated well with the facial morphology measures and the mutation size.

Early in the disease course, before ataxia onset, the prolonged latency of facial motor nerve, jaw jerk and blink reflex suggest the facial nerve involvement^{28,40}. The shrinking of cheek is related to the muscle atrophy and /or loss of fat within the tissue. Axonal damage of cranial nerves appears later during the disease progression.

The strong involvement of cranial nerves in SCA2 has been substantiated previously by histological data. Gierga et al,²⁸ observed significant cell loss and astrogliosis in all studied cases as well as atrophy and myelin loss of associated fibres in most of them. The preferential affection of peripheral nerves such as the facial and trigeminal nerve during the preclinical period is in excellent agreement with other studies of prodromal SCA2, which showed electrophysiological alterations in a pattern of peripheral neuropathy with central pathway damage and neuronal cell loss to depend directly on the CAG repeat expansion size of the ATXN2 gene^{9, 18,40-58}. The physiological function of the Ataxin-2 protein and its highly conserved orthologues until yeast and plants⁵⁹⁻⁶² appears to be the recruitment of fat stores, amino acid reserves in the muscle mass, and glycogen from liver during times of bioenergetics deficit^{31,63-66}, probably through its influence on the endocytosis machinery and on mTOR pathway growth signalling⁶⁷⁻⁷³. To adapt cells to hunger periods, Ataxin-2 performs RNA processing tasks at stress granules and influences mRNA translation⁷⁴⁻⁷⁷, selectively for mitochondrial precursor proteins that are involved in the breakdown of fatty acids, amino acids and pyruvate^{63,77-79}. Indeed, the preferential effects of Ataxin-2 mutations on RNA processing and on mitochondrial factors may become helpful in the diagnosis and progression analysis of SCA2 patients via blood sample RNA sequencing^{80,81}, thus explain a loss of fat tissue and muscle mass may occur through the immobility or cranial neuropathy at late stages of SCA2.

The loss of retroorbital soft tissue and the altered facial expression indeed are known as a very early

feature of SCA2. This is probably due to a loss of fat volume and of muscle mass, given that facial shape in profile photos is known to reflect total fat proportion and to correlate reasonably well with body mass index,⁸² rather than correlating to skeletal parameters [83]. It shows obvious changes with increasing age⁸⁴. Previous investigations of patients with enophthalmos demonstrated that the retro-orbital fat has strong similarities with the buccal fat pad, regarding their admixture with mast cells/endothelial cells/collagen, while truncal adipose tissue is significantly different⁸⁵. This may explain the preferential affection of the face at a SCA2 stage when the body weight is still normal. The results from this study offer a novel tool for the neurophysiological follow up and evaluation of SCA2 patients, allowing to determine the ideal moment for starting a preventive therapy such as hypercaloric high-carbohydrate diets, which were recently reported as promising in the treatment of ALS patients⁸⁶. Such a neuroprotective approach might delay the age at onset or slowing disease progression, but this would be difficult to demonstrate convincingly, unless objective quantitative measurements of disease endophenotypes and electrophysiological deficits can be assessed as well.

This pioneer study can be expanded to an analysis of 3D stereophotogrammetry and geometric morphometrics⁸⁷⁻⁹⁰ as well as to a follow-up longitudinal evaluation, in order to assess the progression of these abnormalities over years within individuals.

As conclusion, these electrophysiological and morphological features offer new insights into the prodromal phenotype of SCA2, while at the same time giving new clues about the role of ATXN2 for muscle atrophy, neuronal energy balance and lipid metabolism. In a clinical setting, our findings could help to the early diagnosis and the design of future therapeutic interventions.

Acknowledgments

We are grateful to the SCA2 patients, preclinical mutation carriers and the control individuals who participated in this study as well as to the Cuban Ministry of Health for their cooperation

References

1. Wadia NH, Swami RK. A new form of hereditary familial spinocerebellar degeneration with slow eye movements (nine families). *Brain* 1971; 94(2):359-74. <https://doi.org/10.1093/brain/94.2.359>.

2. Auburger G, Diaz GO, Capote RF, Sanchez SG, Perez MP, del Cueto ME, et al. Autosomal dominant ataxia: genetic evidence for locus heterogeneity from a Cuban founder-effect population. *Am J Hum Genet* 1990; 46(6): 1163-77. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1683834/>.
3. Hernández A1, Magariño C, Gispert S, Santos N, Lunkes A, Orozco G. Genetic mapping of the spinocerebellar ataxia 2 (SCA2) locus on chromosome 12q23-q24.1. *Genomics* 1995; 25(2):433-5. <http://www.sciencedirect.com/science/article/pii/S088875439580043L>.
4. Gispert S, A Lunkes, N Santos, G Orozco, D Ha-Hao, T Ratzlaff, et al. Localization of the candidate gene D-amino acid oxidase outside the refined I-cM region of spinocerebellar ataxia 2. *Am J Hum Genet* 1995; 57(4): 972-5. <https://www.ncbi.nlm.nih.gov/pmc/?term=Gispert-S+et+al+1993+Nat+Genet>.
5. Pulst SM, Nechiporuk A, Nechiporuk T, Gispert S, Chen XN, Lopes-Cendes I, et al. Moderate expansion of a normally biallelic trinucleotide repeat in spinocerebellar ataxia type 2. *Nat Genet* 1996; 14(3): 269-76. <https://www.ncbi.nlm.nih.gov/pubmed/?term=Pulst-SM+et+al+1996+Nat+Genet>.
6. Sanpei K, Takano H, Igarashi S, Sato T, Oyake M, Sasaki H, et al. Identification of the spinocerebellar ataxia type 2 gene using a direct identification of repeat expansion and cloning technique, DIRECT. *Nat Genet* 1996; 14(3): 277-84. <https://www.ncbi.nlm.nih.gov/pubmed/8896556>
7. Velázquez L. Ataxia Espinocerebelosa tipo 2. Diagnóstico, pronóstico y evolución. 3 ed. La Habana: Editorial Ciencias Médicas 2012. <http://www.bvscuba.sld.cu/libro/ataxia-espinocerebelosa-tipo-2-diagnostico-pronostico-y-evolucion/>.
8. Velázquez-Pérez L, Fernandez-Ruiz J, Díaz R, Pérez-González R, Canales ON, Sánchez CG, et al. Spinocerebellar ataxia type 2 olfactory impairment shows a pattern similar to other major neurodegenerative diseases. *J Neurol* 2006; 253: 1165-9. DOI 10.1007/s00415-006-0183-2.
9. Velázquez PL, Rodríguez LR, Sánchez CG, Laffita MJM, Almaguer ML, Aguilera RR, et al. Comprehensive characterization of spinocerebellar ataxia type 2 in Cuba and its application in intervention projects. *Rev Cub.Salud Pública [serial on the Internet]*. 2011; 37(3): 230-44. [http:// dx. doi. org/10.1590/S0864-34662011000300006](http://dx.doi.org/10.1590/S0864-34662011000300006)
10. Velázquez-Pérez L, Seifried C, Santos-Falcón N, Abele M, Ziemann U, Almaguer LE, et al. Saccade velocity is controlled by polyglutamine size in spinocerebellar ataxia 2. *Ann Neurol* 2004; 56(3): 444-7. <http://onlinelibrary.wiley.com/doi/10.1002/ana.20220/pdf>.
11. Velázquez PL, Seifried C, Abele M, Wirjatijasa F, Rodríguez LR, Santos FN, Sánchez CG, Almaguer ML, et al. Saccade velocity is reduced in presymptomatic spinocerebellar ataxia type 2. *Clin Neurophysiol* 2009; 120(3): 632-5. doi: 10.1016/j.clinph.2008.12.040. <https://www.ncbi.nlm.nih.gov/pubmed/>.
12. Paneque M, Lemos C, Sousa A, Velázquez L, Fleming M, Sequeiros J. Role of the Disease in the Psychological Impact of Pre-Symptomatic Testing for SCA2 and FAP ATTRV30M: Experience with the Disease, Kinship and Gender of the Transmitting Parent. *J Genet Couns* 2009; 18(5): 483-93. doi: 10.1007/s10897-009-9240-1.
13. Rodríguez-Labrada R, Velázquez-Pérez L, Seifried C, Canales-Ochoa N, Auburger G, Medrano-Montero J, et al. Saccadic latency is prolonged in Spinocerebellar Ataxia type 2 and correlates with the frontal-executive dysfunctions. *J Neurol Sci* 2011; 306 (1-2): 103-7. doi: 10.1016/j.jns.2011.03.033.
14. Rodríguez-Labrada R, Velázquez-Pérez L, Auburger G, Ziemann U, Canales-Ochoa N, Medrano-Montero J, et al. Spinocerebellar ataxia type 2: Measures of saccade changes improve power for clinical trials. *Mov Disord* 2016; 31(4):570-8. doi: 10.1002/mds.26532. <http://onlinelibrary.wiley.com/doi/10.1002/mds.26532/epdf>.
15. Almaguer-Mederos LE, Aguilera Rodríguez R, González Zaldivar Y, Almaguer Gotay D, Cuello Almarales D, Laffita Mesa J, et al. Estimation of survival in spinocerebellar ataxia type 2 Cuban patients. *Clin Genet* 2013; 83(3): 293-4. doi: 10.1111/j.1399-0004.2012.01902.x. <http://onlinelibrary.wiley.com/doi/10.1111/j.1399-0004.2012.01902.x/epdf>.
16. Almaguer-Mederos LE, Falcón NS, Almira YR, Zaldivar YG, Almarales DC, Góngora EM, et al. Estimation of the age at onset in spinocerebellar ataxia type 2 Cuban patients by survival analysis. *Clin Genet* 2010; 78(2):169-74. doi: 10.1111/j.1399-0004.2009.01358.x. <http://onlinelibrary.wiley.com/doi/10.1111/j.1399-0004.2009.01358.x/epdf>.

17. Velázquez-Pérez L, Rodríguez-Labrada R, García-Rodríguez JC, Almaguer-Mederos LE, Cruz-Mariño T, Laffita-Mesa JM. A Comprehensive Review of Spinocerebellar Ataxia Type 2 in Cuba. *Cerebellum* 2011; 10(2):184-98. doi: 10.1007/s12311-011-0265-2. <https://link.springer.com/article/10.1007%2Fs12311-011-0265-2>.
18. Rodríguez-Labrada R, Velázquez-Pérez L, Canales ON, Galicia PL, Haro VR, Sanchez CG, et al. Subtle Rapid Eye Movement Sleep Abnormalities in Presymptomatic Spinocerebellar Ataxia Type 2 Gene Carriers. *Mov Disord* 2011; 26(2): 347-50. doi: 10.1002/mds.23409.
19. Auburger GWJ. Spinocerebellar ataxia type 2. In: Subramony SH, Dürr A, editors. *Handbook of Clinical Neurology. Ataxic Disorders*. Vol. 103 (3rd series), 2012, p. 423-36. doi:10.1016/B978-0-444-51892-7.00026-7
20. Della NR, Foresti S, Tessa C, Moretti M, Ginestroni A, Gavazzi C, et al. ADC mapping of neurodegeneration in the brainstem and cerebellum of patients with progressive ataxias. *Neuroimage* 2004; 22(2): 698-705. DOI: 10.1016/j.neuroimage.2004.01.035.
21. Mohit H, Bhalt Richard F, Donad B. Chronic Cerebellar Degeneration. In: *Textbook of Internal Medicine*. William N. Kelley. 2da ed. Lippincott Company (Ed), Philadelphia, New York 1992, pp 2176-7
22. Orozco G, Nodarse A, Cordoves R, Auburger G. Autosomal dominant cerebellar ataxia: Clinical analysis of 263 patients from a homogeneous population in Holguin, Cuba. *Neurology* 1990; 40(9):1369-75.
23. Estrada R, Galarraga J, Orozco G, Nodarse A, Auburger G. Spinocerebellar ataxia 2 (SCA2): Morphometric analyses in 11 autopsies characterize it as an olivo-ponto-cerebellar atrophy (OPCA) plus. *Acta Neuropathol* 1999; 97(3): 306-10. <https://doi.org/10.1007/s004010050989>.
24. Rüb U, Schultz C, Del Tredici K, Gierga K, Reifenberger G, de Vos RA, et al. Anatomically based guidelines for systematic investigation of the central somatosensory system and their application to a spinocerebellar ataxia type 2 (SCA2) patient. *Neuropathol Appl Neurobiol* 2003; 29(5): 418-33. <https://onlinelibrary.wiley.com/doi/epdf/10.1046/j.1365-2990.2003.00504.x>.
25. Rüb U, Bürk K, Schöls L, Brunt ER, de Vos RA, Diaz GO, et al. Damage to the reticulotegmental nucleus of the pons in spinocerebellar ataxia type 1, 2, and 3. *Neurology* 2004; 63(7): 1258-63
26. Rüb U, Gierga K, Brunt ER, de Vos RA, Bauer M, Schöls L, et al. Spinocerebellar ataxias types 2 and 3: degeneration of the pre-cerebellar nuclei isolates the three phylogenetically defined regions of the cerebellum. *J Neural Transm* 2005; 112(11): 1523-45. DOI 10.1007/s00702-005-0287-3.
27. Rüb U, Schöls L, Paulson H, Auburger G, Kermer P, Jen JC, et al. Clinical features, neurogenetics and neuropathology of the polyglutamine spinocerebellar ataxias type 1, 2, 3, 6 and 7. *Prog Neurobiol* 2013; 104: 38-66. doi: 10.1016/j.pneurobio.2013.01.001.
28. Gierga K, Buró K, Bauer M, Orozco DG, Auburger G, Schultz C, et al. Involvement of the cranial nerves and their nuclei in spinocerebellar ataxia type 2 SCA2). *Acta Neuropathol* 2005; 109(6): 617-31. DOI: 10.1007/s00401-005-1014-8
29. Lastres-Becker I, Brodesser S, Lütjohann D, Azizov M, Buchmann J, Hintermann E, et al. Insulin receptor and lipid metabolism pathology in ataxin-2 knock-out mice. *Hum Mol Genet* 2008; 17(10):1465-81. doi: 10.1093/hmg/ddn035.
30. Hoche F, Balikó L, den Dunnen W, Steinecker K, Bartos L, Sáfrány E, et al. Spinocerebellar Ataxia Type 2 (SCA2): Identification of Early Brain Degeneration in One Monozygous Twin in the Initial Disease Stage. *Cerebellum* 2011; 10(2): 245-53. doi: 10.1007/s12311-010-0239-9.
31. Seidel G, Meierhofer D, Şen NE, Guenther A, Krobisch S, Auburger G. Quantitative Global Proteomics of Yeast PBP1 Deletion Mutants and Their Stress Responses Identifies Glucose Metabolism, Mitochondrial, and Stress Granule Changes. *J Proteome Res* 2017; 16(2): 504-15. doi: 10.1021/acs.jproteome.6b00647.
32. Zhang B, Li L, Chen L, Huang J. Clinical manifestations and gene mutation in a case of Machado-Joseph disease. *Neural Regen Res* 2012; 7(35): 2842-7. <http://doi.org/10.3969/j.issn.1673-5374.2012.35.013>
33. Bettencourt C, Lima M. Machado-Joseph Disease: from first descriptions to new perspectives. *Orphanet J Rare Dis* 2011; 6: 35. doi: 10.1186/1750-1172-6-35.
34. Paulson H. Machado-Joseph Disease/Spinocerebellar Ataxia Type 3. *Handb Clin Neurol* 2012; 103: 437-49. doi: 10.1016/B978-0-444-51892-7.00027-9.
35. Trouillas P, Takayanagi T, Hallett M, Currier RD, Subramony SH, Wessel K, et al. International Cooperative Ataxia Rating Scale for

- pharmacological assessment of the cerebellar syndrome. *J Neurol Sci* 1997; 145(2): 205-11. [http://dx.doi.org/10.1016/S0022-510X\(96\)00231-6](http://dx.doi.org/10.1016/S0022-510X(96)00231-6).
36. Kimura J. *Electrodiagnosis in Diseases of Nerve and Muscle: principles and practice*. 3er ed. Oxford university. Press; 2001. ISBN-10: 0195129776. ISBN-13: 9780195129779.
 37. Ward RE. Facial morphology as determined by anthropometry: keeping it simple. *J Craniofac Genet Dev Biol* 1989; 9(1): 45-60.
 38. Mayoral J, Mayoral G. *Principios fundamentales y práctica*. Científico Técnica, La Habana: 1986.
 39. Imbert G, Saudou F, Yvert G, Devys D, Trottier Y, Garnier JM, et al. Cloning of the gene for spinocerebellar ataxia 2 reveals a locus with high sensitivity to expanded CAG/glutamine repeats. *Nat Genet* 1996; 14(3): 285-91. DOI: 10.1038/ng1196-285
 40. Medrano Montero J, Velázquez Pérez LC, Canales Ochoa N, Almaguer Mederos LE. Electrophysiological nerves pattern in patients and asymptomatic carriers with Spinocerebellar Ataxia type 2 (SCA2). *Society Proceedings / Clinical Neurophysiology* 2008; 119; e99–e164. doi:10.1016/j.clinph.2008.04.158
 41. Jacobi H, Hauser TK, Giunti P, Globas C, Bauer P, Schmitz-Hübsch T, et al. Spinocerebellar ataxia types 1, 2, 3 and 6: the Clinical spectrum of ataxia and morphometric brainstem and cerebellar findings. *Cerebellum* 2012; 11(1): 155-66. doi: 10.1007/s12311-011-0292-z.
 42. Klockgether T. Ataxias. In: Goetz CG editors. *Textbook of Clinical Neurology*. 3rd ed, Philadelphia: Saunders an imprint of Elsevier Inc (Ed); 2007. ISBN: 9781416036180.
 43. Geiner S, Horn AK, Wadia NH, Sakai H, Büttner-Ennever JA. The neuroanatomical basis of slow saccades in spinocerebellar ataxia type 2 (Wadia-subtype). *Prog Brain Res* 2008; 171: 575–81. doi: 10.1016/S0079-6123(08)00683-3.
 44. Velázquez-Pérez L, Rodríguez-Labrada R, Canales-Ochoa N, Montero JM, Sánchez-Cruz G, Aguilera-Rodríguez R, et al. Progression of early features of spinocerebellar ataxia type 2 in individuals at risk: a longitudinal study. *Lancet Neurol* 2014; 13 (5): 482-9. [http://dx.doi.org/10.1016/S1474-4422\(14\)70027-4](http://dx.doi.org/10.1016/S1474-4422(14)70027-4).
 45. Velázquez-Pérez L, Rodríguez-Labrada R, Torres-Vega R, Medrano-Montero J, Vázquez-Mojena Y, Auburger G, et al. Abnormal corticospinal tract function and motor cortex excitability in non-ataxic SCA2 mutation carriers: A TMS study. *Clin Neurophysiol* 2016; 127(8): 2713-9. doi: 10.1016/j.clinph.2016.05.003.
 46. Velázquez-Pérez L, Rodríguez-Labrada R, Torres-Vega R, Montero JM, Vázquez-Mojena Y, Auburger G, et al. Central motor conduction time as prodromal biomarker in spinocerebellar ataxia type 2. *Mov Disord* 2016; 31(4):603-4. doi: 10.1002/mds.26555.
 47. Velázquez-Pérez L, Tünnerhoff J, Rodríguez-Labrada R, Torres-Vega R, Belardinelli P, Medrano-Montero J, et al. Corticomuscular Coherence: a Novel Tool to Assess the Pyramidal Tract Dysfunction in Spinocerebellar Ataxia Type 2. *Cerebellum* 2017; 16(2): 602-6. DOI 10.1007/s12311-016-0827-4
 48. Velázquez-Pérez L, Sanchez Cruz G, Canales Ochoa N, et al. Electrophysiological features in patients and presymptomatic relatives with spinocerebellar ataxia type 2. *J Neurol Sci* 2007; 263:158-64. <http://www.pubpdf.com/pub/17706249/Electrophysiological-features-in-patients-and-presymptomatic-relatives-with-spinocerebellar-ataxia-t>.
 49. Halbach-MV, Gispert S, Stehning T, Damrath E, Walter M, Auburger G. Atxn2 Knockout and CAG42-Knock-in Cerebellum Shows Similarly Dysregulated Expression in Calcium Homeostasis Pathway. *Cerebellum* 2017; 16(1): 68-81. doi: 10.1007/s12311-016-0762-4. <https://www.ncbi.nlm.nih.gov/labs/articles/26868665/>.
 50. Schöls L, Reimold M, Seidel K, Globas C, Brockmann K, Hauser TK, et al. No parkinsonism in SCA2 and SCA3 despite severe neurodegeneration of the dopaminergic substantia nigra. *Brain* 2015; 138(Pt 11):3316-26. doi: 10.1093/brain/awv255.
 51. Schöls L, Gispert S, Vorgerd M, Menezes Vieira-Saecker AM, Blanke P, Auburger G, et al. Spinocerebellar ataxia type 2. Genotype and phenotype in German kindreds. *Arch Neurol* 1997; 54(9):1073-80. doi:10.1001/archneur.1997.00550210011007.
 52. Damrath E, Heck MV, Gispert S, Azizov M, Nowock J, Seifried C, et al. ATXN2-CAG42 Sequesters PABPC1 Into Insolubility and Induces FBXW8 in Cerebellum of Old Ataxic Knock-In Mice. *PLoS Genet* 2012; 8(8):e1002920. doi: 10.1371/journal.pgen.1002920.
 53. Riess O, Laccione FA, Gispert S, Schöls L, Zühlke C, Vieira-Saecker AM, et al. SCA2 trinucleotide expansion in German SCA patients.

- Neurogenetics 1997; 1(1): 59-64. <https://www.ncbi.nlm.nih.gov/pubmed/10735276>.
54. Elden AC, Kim HJ, Hart MP, Chen-Plotkin AS, Johnson BS, Fang X, et al. Ataxin-2 intermediate-length polyglutamine expansions are associated with increased risk for ALS. *Nature* 2010; 26; 466(7310): 1069-75. doi: 10.1038/nature09320. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2965417/>.
 55. Lee T, Li YR, Ingre C, Weber M, Grehl T, Gredal O, et al. Ataxin-2 intermediate-length polyglutamine expansions in European ALS patients. *Hum Mol Genet* 2011; 20(9): 1697-700. doi: 10.1093/hmg/ddr045.
 56. Gispert S, Kurz A, Waibel S, Bauer P, Liepelt I, Geisen C, et al. The modulation of Amyotrophic Lateral Sclerosis risk by ataxin-2 intermediate polyglutamine expansions is a specific effect. *Neurobiol Dis* 2012; 45(1): 356-61. doi: 10.1016/j.nbd.2011.08.021.
 57. Lahut S, Ömür Ö, Uyan Ö, Ağım ZS, Özoğuz A, Parman Y, et al. ATXN2 and its neighbouring gene SH2B3 are associated with increased ALS risk in the Turkish population. *PLoS One* 2012; 7(8): e42956. doi: 10.1371/journal.pone.0042956.
 58. Dansithong W, Paul S, Figueroa KP, Rinehart MD, Wiest S, Pflieger LT, et al. Ataxin-2 regulates RGS8 translation in a new BAC-SCA2 transgenic mouse model. *PLoS Genet* 2015; 11(4): e1005182. doi: 10.1371/journal.pgen.1005182.
 59. Jiménez-López D, Guzmán P. Insights into the evolution and domain structure of Ataxin-2 proteins across eukaryotes. *BMC Res Notes* 2014; 7: 453. doi: 10.1186/1756-0500-7-453. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4105795/>.
 60. Jiménez-López D, Bravo J, Guzmán P. Evolutionary history exposes radical diversification among classes of interaction partners of the MLL domain of plant poly(A)-binding proteins. *BMC Evol Biol* 2015; 15: 195. doi: 10.1186/s12862-015-0475-1. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4574140/>.
 61. Ralser M, Albrecht M, Nonhoff U, Lengauer T, Lehrach H, Krobitsch S. An integrative approach to gain insights into the cellular function of human ataxin-2. *J Mol Biol* 2005; 346(1): 203-14. DOI: 10.1016/j.jmb.2004.11.024
 62. Satterfield TF, Jackson SM, Pallanck LJ. A *Drosophila* homolog of the polyglutamine disease gene SCA2 is a dosage-sensitive regulator of actin filament formation. *Genetics* 2002; 162(4): 1687-702. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1462369/pdf/12524342.pdf>.
 63. Meierhofer D, Halbach M, Şen NE, Gispert S, Auburger G. Ataxin-2 (Atxn2)-Knock-Out Mice Show Branched Chain Amino Acids and Fatty Acids Pathway Alterations. *Mol Cell Proteomics* 2016; 15(5): 1728-39. doi: 10.1074/mcp.M115.056770. <http://www.mcponline.org/content/15/5/1728.long>.
 64. Auburger G, Gispert S, Lahut S, Omür O, Damrath E, Heck M, et al. 12q24 locus association with type 1 diabetes: SH2B3 or ATXN2? *World J Diabetes* 2014; 5(3): 316-27. doi: 10.4239/wjd.v5.i3.316. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4058736/>.
 65. Lastres-Becker I, Rüb U, Auburger G. Spinocerebellar ataxia 2 (SCA2). *Cerebellum* 2008; 7(2): 115-24. doi: 10.1007/s12311-008-0019-y.
 66. Kiehl TR, Nechiporuk A, Figueroa KP, Keating MT, Huynh DP, Pulst SM. Generation and characterization of Sca2 (ataxin-2) knockout mice. *Biochem Biophys Res Commun* 2006; 339(1):17-24. DOI: 10.1016/j.bbrc.2005.10.186
 67. Lastres-Becker I, Nonis D, Eich F, Klinkenberg M, Gorospe M, Kötter P, et al. Mammalian ataxin-2 modulates translation control at the pre-initiation complex via PI3K/mTOR and is induced by starvation. *Biochim Biophys Acta* 2016; 1862(9): 1558-69. doi: 10.1016/j.bbadis.2016.05.017.
 68. Drost J, Nonis D, Eich F, Leske O, Damrath E, Brunt ER, et al. Ataxin-2 modulates the levels of Grb2 and SRC but not ras signaling. *J Mol Neurosci* 2013; 51(1): 68-81. doi: 10.1007/s12031-012-9949-4. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3739869/>
 69. Nonis D, Schmidt MH, van de Loo S, Eich F, Dikic I, Nowock J, et al. Ataxin-2 associates with the endocytosis complex and affects EGF receptor trafficking. *Cell Signal* 2008; 20(10):1725-39. doi: 10.1016/j.cellsig.2008.05.018.
 70. Ralser M, Nonhoff U, Albrecht M, Lengauer T, Wanker EE, Lehrach H, et al. Ataxin-2 and huntingtin interact with endophilin-A complexes to function in plastin-associated pathways. *Hum Mol Genet* 2005; 14(19): 2893-909. DOI: 10.1093/hmg/ddi321
 71. Takahara T, Maeda T. Transient sequestration of TORC1 into stress granules during heat

- stress. *Mol Cell* 2012; 47(2): 242-52. doi: 10.1016/j.molcel.2012.05.019.
72. DeMille D, Badal BD, Evans JB, Mathis AD, Anderson JF, Grose JH. PAS kinase is activated by direct SNF1-dependent phosphorylation and mediates inhibition of TORC1 through the phosphorylation and activation of Pbp1. *Mol Biol Cell* 2015; 26(3): 569-82. doi: 10.1091/mbc.E14-06-1088. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4310746/>.
 73. Bar DZ, Charar C, Dorfman J, Yadid T, Tafforeau L, Lafontaine DL, et al. Cell size and fat content of dietary-restricted *Caenorhabditis elegans* are regulated by ATX-2, an mTOR repressor. *Proc Natl Acad Sci U S A* 2016; 113(32): E4620-9. doi: 10.1073/pnas.1512156113.
 74. Fittschen M, Lastres-Becker I, Halbach MV, Damrath E, Gispert S, Azizov M, et al. Genetic ablation of ataxin-2 increases several global translation factors in their transcript abundance but decreases translation rate. *Neurogenetics* 2015; 16(3): 181-92. doi: 10.1007/s10048-015-0441-5. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4475250/>.
 75. van de Loo S, Eich F, Nonis D, Auburger G, Nowock J. Ataxin-2 associates with rough endoplasmic reticulum. *Exp Neurol* 2009; 215(1): 110-8. doi: 10.1016/j.expneurol.2008.09.020.
 76. Swisher KD, Parker R. Localization to, and effects of Pbp1, Pbp4, Lsm12, Dhh1, and Pab1 on stress granules in *Saccharomyces cerevisiae*. *PLoS One* 2010; 5(4): e10006. doi: 10.1371/journal.pone.0010006. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2848848/>.
 77. Nonhoff U, Ralser M, Welzel F, Piccini I, Balzeireit D, Yaspo ML, et al. Ataxin-2 interacts with the DEAD/H-box RNA helicase DDX6 and interferes with P-bodies and stress granules. *Mol Biol Cell* 2007; 18(4): 1385-96. DOI: 10.1091/mbc.E06-12-1120. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1838996/>.
 78. Seidel K, Siswanto S, Fredrich M, Bouzrou M, den Dunnen WF, Özerden I, et al. On the Distribution of Intranuclear and Cytoplasmic Aggregates in the Brainstem of Patients with Spinocerebellar Ataxia Type 2 and 3. *Brain Pathol* 2017; 27(3): 345-55. doi: 10.1111/bpa.12412.
 79. Wang X, Chen XJ. A Cytosolic Network Suppressing Mitochondria-Mediated Proteostatic Stress and Cell Death. *Nature* 2015; 524(7566): 481-4. doi: 10.1038/nature14859. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4582408/>.
 80. Vianna MC, Poletto DC, Gomes PF, Valente V, Paçó-Larson ML. *Drosophila* ataxin-2 gene encodes two differentially expressed isoforms and its function in larval fat body is crucial for development of peripheral tissues. *FEBS Open Bio* 2016; 6(11): 1040-53. eCollection 2016. DOI:10.1002/2211-5463.12124. <http://onlinelibrary.wiley.com/doi/10.1002/2211-5463.12124/epdf>.
 81. Sen NE, Drost J, Gispert S, Torres-Odio S, Damrath E, Klinkenberg M, et al. Search for SCA2 blood RNA biomarkers highlights Ataxin-2 as strong modifier of the mitochondrial factor PINK1 levels. *Neurobiol Dis* 2016; 96: 115-26. doi: 10.1016/j.nbd.2016.09.002.
 82. Mayer C, Windhager S, Schaefer K, Mittröcker P. BMI and WHR Are Reflected in Female Facial Shape and Texture: A Geometric Morphometric Image Analysis. *PLoS One* 2017; 12(1): e0169336. doi: 10.1371/journal.pone.0169336.
 83. Krey KF, Dannhauer KH. Morphometric analysis of facial profile in adults. *J Orofac Orthop* 2008; 69(6): 424-36. doi: 10.1007/s00056-008-8803-3.
 84. Halazonetis DJ. Morphometric correlation between facial soft-tissue profile shape and skeletal pattern in children and adolescents. *Am J Orthod Dentofacial Orthop* 2007; 132(4): 450-7. DOI: 10.1016/j.ajodo.2005.10.033
 85. Ilankovan V, Soames JV. Morphometric analysis of orbital, buccal and subcutaneous fats: their potential in the treatment of enophthalmos. *Br J Oral Maxillofac Surg* 1995; 33(1): 40-2.
 86. Wills AM, Hubbard J, Macklin EA, Glass J, Tandan R, Simpson EP, et al. Hypercaloric enteral nutrition in patients with amyotrophic lateral sclerosis: a randomised, double-blind, placebo-controlled phase 2 trial. *Lancet* 2014; 383(9934): 2065-72. doi: 10.1016/S0140-6736(14)60222-1. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4176708/>.
 87. Weinberg SM, Parsons TE, Marazita ML, Maher BS. Heritability of Face Shape in Twins: A Preliminary Study using 3D Stereophotogrammetry and Geometric Morphometrics. *Dent* 2013; 3000 1(1). pii: 14.
 88. Liang S, Wu J, Weinberg SM, Shapiro LG. Improved detection of landmarks on 3D human

- face data. Conf Proc IEEE Eng Med Biol Soc 2013; 6482-5. doi: 10.1109/EMBC.2013.6611039.
89. Kustár A, Forró L, Kalina I, Fazekas F, Honti S, Makra S, et al. FACE-R--a 3D database of 400 living individuals' full head CT- and face scans and preliminary GMM analysis for craniofacial reconstruction. *J Forensic Sci* 2013; 58(6): 1420-8. doi: 10.1111/1556-4029.12215. <http://onlinelibrary.wiley.com/doi/10.1111/1556-4029.12215/pdf>.
90. Wellens HL, Kuijpers-Jagtman AM, Halazonetis DJ. Geometric morphometric analysis of craniofacial variation, ontogeny and modularity in a cross-sectional sample of modern humans. *J Anat* 2013; 222(4): 397-409. doi: 10.1111/joa.12027. <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC3610033/pdf/joa0222-0397.pdf>